RESEARCH ARTICLE

Open Access

Effect of pasak bumi (*Eurycoma longifolia Jack*), DHA, and seluang fish (*Rasbora* spp.) on neuroinflammation and neurotransmitter alterations in malnourished rats



Triawanti^{1*}, Didik Dwi Sanyoto², Meitria Syahadatina Noor³, Dimas Ikhsan Airlangga⁴

ABSTRACT

Background: Malnutrition has detrimental effects on brain development, leading to neuroinflammatory and neurotransmitter disorders. A nutrient-rich diet is advocated to mitigate these effects. In South Kalimantan, natural resources such as pasak bumi (*Eurycoma longifolia* Jack.) and seluang fish (*Rasbora* spp.) are recognized for their potential antioxidant and anti-inflammatory properties.

Objective: This study aimed to evaluate the effects of pasak bumi in comparison with seluang fish and DHA on neuroinflammation and neurotransmitter imbalances in malnourished rats.

Methods: The study involved dividing malnourished rats into six experimental groups: (1) untreated control, (2) treated with pasak bumi extract, (3) treated with DHA, (4) treated with a combination of DHA and pasak bumi extract, (5) treated with seluang fish, (6) treated with a combination of seluang fish and pasak bumi extract; and a control group of normal rats receiving standard feed and placebo. The primary outcomes measured were levels of IL-6, TNF- α , and serotonin.

Results: The study revealed that malnutrition in rats significantly elevated IL-6 and TNF- α levels. Treatments involving pasak bumi extract, both alone and in combination with DHA or seluang fish, reduced IL-6 levels. Similarly, combination of pasak bumi extract and DHA or seluang fish lowered TNF- α levels than single treatment of pasak bumi extract (p > 0.05). All treatments did not reduce the serotonin level.

Conclusion: The findings of this study underscore the potent anti-inflammatory capabilities of pasak bumi extract, particularly when combined with DHA or seluang fish, in mitigating the inflammatory response in malnourished rats.

Keywords: malnutrition, pasak bumi, seluang fish, neuroinflammation, neurotransmitter

Introduction

Malnutrition has long been a significant public health concern in Indonesia. The latest data from the *Studi Status Gizi Indonesia* (SSGI) in 2021 indicates a decrease in stunting prevalence from 27.7% to 24.4% compared to the previous year. However, this persistent issue continues to affect various regions unevenly. Notably, South Kalimantan ranks sixth nationally in stunting among children under five and third in the underweight category, which is particularly alarming [1].

Protein-deficient malnutrition can disrupt the synthesis of enzymes that function as antioxidants, reducing their numbers [2,3]. This results in an imbalance between antioxidants and oxidative stress in the brain, causing excess free radicals [4]. These free radicals can damage cellular components, including proteins, DNA, membrane phospholipids, and enzymes [5,6]. Furthermore, malnutrition disrupts the immune system's balance, reducing anti-inflammatory biomarkers and increasing proinflammatory ones [7,8]. This harmful alteration

¹Department of Biochemistry and Biomolecular, Medical Faculty, Lambung Mangkurat University, Banjarmasin, 70232, Indonesia ²Department of Biomedic, Anatomy Division, Medical Faculty, Lambung Mangkurat University, Banjarmasin, 70232, Indonesia

³Department of Public Health, Medical Faculty, Lambung Mangkurat University, Banjarmasin, 70232, Indonesia

⁴Undergraduate Medical Faculty, Brawijaya University, Malang, 65145, Indonesia

^{*}Corresponding author: Jl. Veteran No 128 Banjarmasin 70232. Email: triawanti@ulm.ac.id

in the immune system is suggested to contribute to neuroinflammation in the brain, potentially impairing neuronal development and affecting a child's intelligence over time [9,10].

One approach to overcome malnutrition is providing highly nutritious, particularly high-protein foods, to protein-malnourished children [11,12]. A previous study demonstrated that the administration of seluang fish, high in protein and commonly consumed in South Kalimantan, increased IGF-1 levels, benefiting bone growth and haemoglobin levels in an animal model [13]. This intervention also alleviated oxidative stress in the brain [14] and improved memory in malnourished rats [15]. Another local nutrient source, pasak bumi (Eurycoma longifolia Jack), is believed to hold the potential for addressing brain damage caused by malnutritioninduced inflammation [8-17]. The active compounds in E. longifolia, such as quassinoids, biphenylneolignan, tirucallane-type triterpenes, β-carboline alkaloids, canthin-6-one alkaloids, and squalene derivatives, have demonstrated antioxidant and anti-inflammatory properties in previous studies [18,19].

This study employed an experimental rat model (Rattus norvegicus) with protein deficiency. First, we focused on determining the effective dose of pasak bumi extract to mitigate the adverse effects of malnutrition [19]. The results indicated that rats given a 15 mg/kg BW dose of pasak bumi extract showed improvements in oxidative stress and higher retention of spatial memory compared to the control group and other doses [19]. Second, we compared this effective dose with the administration of seluang fish, DHA, and the combination of each with pasak bumi extract in overcoming neuroinflammation and neurotransmitter disorders in the brains of malnourished rats. We assessed neuroinflammation through IL-6 and TNF- α levels and evaluated neurotransmitter function using serotonin levels [20,21].

Methods

Ethical approval

This study received ethical approval from the Health Research Ethics Commission, Faculty of Medicine, Lambung Mangkurat University (approval No. 061/KEPK-FK UNLAM/EC/II/2020).

Feed composition

In this study, we used two distinct types of feed: a low-protein feed and a standard feed. The low-protein feed is comprised of the AIN-76A purified rodent diet [Dyets Inc., USA], which includes cornstarch (183 g/kg), corn oil (50 g/kg), cellulose (50 g/kg), DL-methionine (0.9 g/kg), sucrose (609.1 g/kg), casein (60 g/kg), a mineral mix#200000 (35 g/kg), a vitamin mix #300050 (10 g/kg), and choline bitartrate (2 g/kg). Conversely, the standard feed contains 20–22% protein, 5–7% fat, 5–7% cinder, 3–5% fiber, 9–11% calcium, 0.6–0.8% phosphorus, and an energy content of 2900–3100 kcal.

Preparation of experimental animal malnutrition model

Newborn rats were breastfed by mothers on a low-protein diet (AIN76A, 6% protein) for four weeks, then continued on the same diet post-weaning for another four weeks. Rats with serum protein levels below 4.7 g/dL were classified as malnourished. Serum protein was measured from 1 mL of blood obtained via tail vein puncture and centrifugation [22].

Treatments

Rats were divided into seven groups (five rats per group):

- Normal control: healthy rats on standard feed and placebo
- Untreated control: malnourished rats on standard feed and placebo
- P1: malnourished rats on standard feed and pasak bumi ethanol extract (15 mg/kg BW)
- P2: malnourished rats on standard feed and DHA (1 mg/kg BW)
- P3: malnourished rats on standard feed and combination DHA (1 mg/kg BW) and pasak bumi extract (15 mg/kg BW)

- P4: malnourished rats on seluang fish
- P5: malnourished rats on combonation seluang fish and pasak bumi extract (15 mg/kg BW).

The treatment duration was five weeks, based on prior studies identifying the optimal antioxidant activity dose [19].

Examination of TNF-α, IL-6, and serotonin levels

At the 5-week mark, rats were euthanized using anaesthetic drug and cardiac puncture, followed by brain extraction for TNF- α , IL-6, and serotonin level measurement via the ELISA method [23]. The procedure was conducted according to the manufacturer's guidelines (BioTechnology Laboratory®).

Data analysis

Data were analyzed using SPSS software. The Shapiro-Wilk test checked data normality. For normally distributed data, an ANOVA test with a 95% confidence level and LSD posthoc test were used. For non-normally distributed data, the non-parametric Kruskal–Wallis test followed by the Mann–Whitney test was applied, maintaining a 95% confidence level.

Results

Effect treatments to IL-6 level

The study demonstrates that malnourished rats, serving as the untreated control group, exhibited elevated IL-6 levels in the brain compared to a normal group (Figure 1). This indicates that malnutrition may increase the inflammatory response. When malnourished rats were treated with pasak bumi extract, their IL-6 levels were lower than those in the untreated group, suggesting the extract's potential to mitigate IL-6 levels. Conversely, treatment with DHA alone did not significantly alter IL-6 levels compared to the untreated control. However, a combined treatment of pasak bumi extract and DHA significantly reduced IL-6 levels, surpassing DHA treatment alone.

Similarly, administering seluang fish alone did not impact IL-6 levels, maintaining them on par

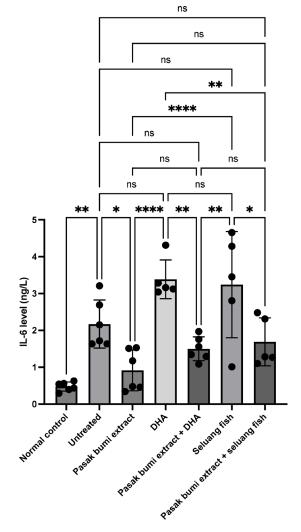


Figure 1. IL-6 levels of malnourished rat brain after intervention. Normal control: healthy rats on standard feed and placebo; Untreated control: malnourished rats on standard feed and placebo; P1: malnourished rats on standard feed and pasak bumi extract (15 mg/kg BW); P2: malnourished rats on standard feed and DHA (1 mg/kg BW); P3: malnourished rats on standard feed, pasak bumi extract (15 mg/kg BW) and DHA (1 mg/kg BW); P4: malnourished rats on seluang fish; P5: malnourished rats on pasak bumi extract (15 mg/kg BW) and seluang fish

with the untreated group. Notably, a combined treatment involving pasak bumi extract and seluang fish effectively reduced IL-6 levels. These findings underscore the crucial role of pasak bumi extract in diminishing IL-6 levels in malnourished rats.

Effect treatments to TNF-alpha levels

Our findings indicate that TNF- α levels were elevated in the malnourished, untreated group compared to the normal group (Figure 2). Notably, malnourished rats treated with a combination

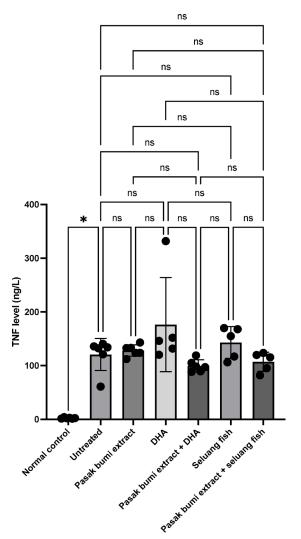


Figure 2. TNF-α levels of malnourished rat brain after intervention Normal control: healthy rats on standard feed and placebo; Untreated control: malnourished rats on standard feed and placebo; P1: malnourished rats on standard feed and pasak bumi extract (15 mg/kg BW); P2: malnourished rats on standard feed and DHA (1 mg/kg BW); P3: malnourished rats on standard feed, pasak bumi extract (15 mg/kg BW), and DHA (1 mg/kg BW); P4: malnourished rats on seluang fish; P5: malnourished rats on pasak bumi extract (15 mg/kg BW) and seluang fish

of pasak bumi extract and DHA exhibited lower TNF- α levels than those in the untreated group. Furthermore, this combination was more effective in reducing TNF- α levels in malnourished rats than treatments involving only pasak bumi extract, DHA, or seluang fish.

Effect treatments to serotonin level

Regarding serotonin levels, the data revealed that the average brain serotonin in malnourished rats exceeded that of normal rats (Figure 3).

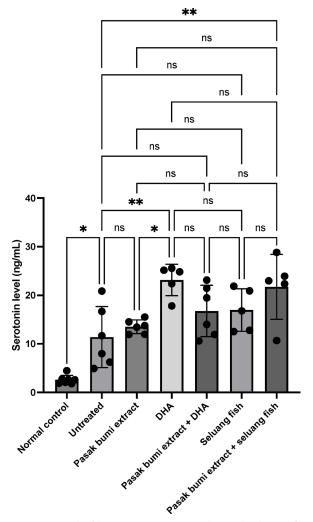


Figure 3. Level of brain serotonin in malnourished rats after intervention Normal control: healthy rats on standard feed and placebo; Untreated control: malnourished rats on standard feed and placebo; P1: malnourished rats on standard feed and pasak bumi extract (15 mg/kg BW); P2: malnourished rats on standard feed and DHA (1 mg/kg BW); P3: malnourished rats on standard feed, pasak bumi extract (15 mg/kg BW), and DHA (1 mg/kg BW); P4: malnourished rats on seluang fish; P5: malnourished rats on pasak bumi extract (15 mg/kg BW) and seluang fish

Interventions involving pasak bumi extract, DHA, and seluang fish in malnourished rats resulted in higher serotonin levels compared to the normal control group. Among the intervention groups, treatment with pasak bumi extract alone resulted in the lowest brain serotonin levels.

Discussion

In this study, the effects of seluang fish and DHA on neuroinflammation and neurotransmitter levels in malnourished rats were examined, both as a single

treatment and in combination with pasak bumi extract. The findings revealed that malnourished rats exhibited significantly elevated levels of IL-6 and TNF- α (Figures 1 and 2) compared to the normal group. This aligns with prior research indicating that protein deficiency can increase inflammatory mediators IL-6 and TNF- α levels. Notably, administering pasak bumi extract led to the most substantial reduction in IL-6 levels among all intervention groups. When pasak bumi extract was combined with DHA, IL-6 levels did not differ significantly from those observed with the administration of pasak bumi extract alone. A similar trend was observed with the combination of pasak bumi extract and seluang fish, where IL-6 levels were not significantly different from those in rats treated solely with pasak bumi extract. Furthermore, both the combination of pasak bumi extract with DHA and the combination with seluang fish resulted in lower TNF- α levels compared to their respective treatments alone. These results suggest that pasak bumi extract possesses considerable anti-inflammatory potential.

Pasak bumi extract demonstrates significant anti-inflammatory effects, as evidenced by various studies, through mechanisms such as (i) inhibiting the release of inducible nitric oxide synthase (iNOS), nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB), and interleukin-6 (IL-6), which are critical inflammation-related proteins, within the lipopolysaccharide (LPS)induced NF-κB signaling pathway [24]; and (ii) suppressing the activity of nitric oxide (NO), iNOS, and cyclooxygenase-2 (COX2), while also protecting against death in LPS-induced septic shock models in mice [25,26]. Adding docosahexaenoic acid (DHA) and fish to the treatment regimen enhances the anti-inflammatory potential of the intervention.

DHA, an omega-3 fatty acid, serves as an anti-inflammatory mediator [27,28], impacting inflammation through various mechanisms related to the composition of cell membranes [29]. Alterations in membrane composition can influence fluidity and cell signaling, thereby affecting gene expression and the production of

lipid mediators. Inflammation-involved cells typically contain high levels of arachidonic acid, an omega-6 fatty acid [30]. However, oral supplementation with EPA (eicosapentaenoic acid) and DHA can modify the levels of arachidonic acid, EPA, and DHA. Eicosanoids derived from arachidonic acid, including prostaglandins, thromboxanes, and leukotrienes, play roles in inflammation regulation through processes such as vasoconstriction and enhancing immune responses, often promoting pro-inflammatory mediators [30].

In contrast, EPA produces eicosanoids with antiinflammatory properties, distinct from those derived from arachidonic acid [32]. Both EPA and DHA inhibit additional inflammatory mechanisms, such as neutrophile chemotaxis [33], the production of eicosanoids (like prostaglandins and leukotrienes) from arachidonic acid [34], and the production of cytokines and T-cell reactivity [35]. These effects are mediated through various mechanisms, including activating cell surface molecules (GPR120) and intracellular receptors (PPAR-gamma) that regulate inflammatory cell signaling and gene expression patterns [32,36].

Previous study has also demonstrated that EPA and DHA can inhibit IL-6 and IL-8 production stimulated by endotoxins in human endothelial cell cultures and reduce TNF-alpha production in monocyte cultures induced by endotoxins [32]. Furthermore, EPA reduces the activation of NF-κB triggered by endotoxins in monocytes, correlated with decreased phosphorylation of IκB [38]. The omega-3 fatty acids from marine fish have been studied for their role in reducing inflammation by inhibiting the transcription factor NF-κB, through mechanisms including reducing eicosanoid mediator production from arachidonic acid, increasing production of anti-inflammatory eicosanoids from EPA, enhancing the production of anti-inflammatory and resolvin molecules from EPA and DHA, decreasing leukocyte chemotaxis, suppressing the expression of leukocyte and endothelial adhesion molecules to reduce adhesive interactions, and inhibiting the production of proinflammatory cytokines and proteins by affecting the NF-κB signaling pathway [27].

This study also used seluang fish to treat brain disorders due to malnutrition, one of which was neuroinflammation. Besides containing DHA, seluang fish contains complete amino acids [39]. The group with the combination of pasak bumi extract and seluang fish showed lower levels of IL-6 and TNF- α compared to those given pasak bumi extract or seluang fish alone, and similar to being given a combination of DHA and pasak bumi extract (Figures 2 and 3). Amino acids found in seluang fish include essential and non-essential amino acids. Several studies have proven the role of amino acids in the inflammatory process [40,41,42]. In inflammatory bowel disease (IBD), the anti-inflammatory activity of the following amino acids has been demonstrated: tryptophan, glycine, methionine, cysteine, and arginine [43]. Tryptophan inhibits the activation of proinflammatory cytokines (TNF- α , IL-6, IFN- γ , IL-12p40, IL-1 β , and ICAM-1) [44], glutamine reduces the expression of CCR9, LFA-1, and PSGL-1 produced by Th cells [45], glutamate inhibits T cell response and inflammation [46], cysteine inhibits the expression of local inflammatory mediators (TNF-α, IL-6, IL-12p40, and IL-1β) [47], histidine reduces the production of TNF-α, IL-6, and also inhibits NF-κB signaling [48], arginine reduces proinflammatory cytokines and chemokine expression [49], glycine prevents the increase of IL-1 β and TNF- α [48], taurine inhibits NF-κB activity [50].

Branched-chain amino acids (BCCAs) such as leucine also promote anti-inflammatory effects [51,52,53]. BCAAs modulate inflammation by synthesizing glutamine, which occurs via BCAA transaminases and produces glutamate from α -ketoglutarate [54]. Glutamate is then converted to glutamine using glutamine synthetase [55]. Glutamine affects the release of TNF-α, IL-2, IL-6, IL-10, and IFN-γ [56]. Studies have also shown that glutamine plays a crucial role in the NF-κB signal transduction pathway, contributing to the reduction of local inflammation [57,58]. Glutamine is also involved in the inflammatory pathway in ulcerative colitis [59,60]. Glutamine inhibits this process by increasing the heat shock protein activity, which can reduce the expression of the transcription

factor NF- κ B and stimulate genes associated with the inflammatory response [61]. Administration of short-term glutamine supplementation proved to have a significant decrease in NF- κ B [62, 63]. Glutamine can also reduce inflammation by inhibiting the activation of STAT protein and normalizing the production of nitric oxide [63].

Figure 3 illustrates that serotonin levels in the malnourished group were notably higher than those in the normal group, aligning with existing research indicating an increase in serotonin levels under malnutrition conditions [64,65]. This rise is attributed to the impact of undernutrition, occurring during pre- and post-natal stages, on the serotonin neurotransmission system. Such nutritional deficits activate the brain's serotonergic biosynthetic system and metabolism, consequently elevating the free fraction of L-Tryptophan (FFT), a precursor to serotonin [66,67]. Additionally, protein deficiency alters the plasma albumin's capacity to bind L-Trp, further increasing FFT levels in the plasma. This elevated FFT can cross the blood-brain barrier, which is absorbed by brainstem serotonergic neurons, triggering serotonin synthesis [68]. The unusually high serotonin levels in malnutrition may also result from brain developmental reprogramming due to nutritional deficiencies [69].

Abnormally low serotonin levels can impair learning abilities and memory [70], whereas excessively high levels are similarly detrimental [71]. Elevated serotonin has been linked to the dysregulation of food intake, indicating hyperphagia as a result of fetal programming [72]. Furthermore, high serotonin levels are associated with decreased appetite, increased impulsivity, and diminished reference memory [73]. This study found that all malnourished groups—whether treated with pasak bumi extract, DHA, seluang fish individually, or in combination—exhibited elevated serotonin levels compared to the normal group. Notably, the group treated with pasak bumi extract alone displayed the lowest serotonin levels among the intervention groups, suggesting that pasak bumi extract may prevent serotonin levels from rising excessively.

Deficiency in omega-3 fatty acids leads to increased basal levels and diminished stimulated levels of serotonin [73]. In contrast, serotonin release can diminish under pharmacological stimulation. The restoration of serotonin release impaired by dietary deficiency is achievable through an adequate omega-3 diet during critical neurodevelopmental stages, including at birth, between 7-14 days postnatal, or during lactation [74]. Administering an adequate diet during lactation can replenish the fatty acid composition in the brain and facilitate serotonin release [74]. However, when such a diet is introduced post-weaning, it fails to restore serotonin levels despite normalizing DHA levels within the hippocampal membrane. This finding clarifies why, in the present study, diets rich in DHA and seluang fish did not bring serotonin levels back to normal level, given that the interventions were implemented after the weaning phase. Consequently, it underscores the importance of ensuring an adequate intake of nutrients during the first 1000 days of life to support optimal neurodevelopment and prevent long-term deficits in serotonin levels.

Conclusion

Protein deficiency-induced malnutrition adversely impacts the brain, escalating the neuroinflammatory response and triggering excessive serotonin production. The study reveals that combining pasak bumi extract with DHA, as well as pasak bumi extract with seluang fish, effectively reduces levels of IL-6 and TNF- α in the brains of malnourished rats. Furthermore, pasak bumi extract alone demonstrates a unique ability to diminish serotonin levels in the brains of these rats compared to other interventions. Consequently, the synergistic effects of pasak bumi extract with either DHA or seluang fish highlight their potential as new therapeutic agents for mitigating malnutrition.

Acknowledgment

We thank the DRPM Ministry of Research, Technology/BRIN Republik Indonesia for the total financial support by research grant (contract letter No. 031/E4.1/AK.04.PT/2021) and all people for their best contribution.

Author contributions

Conceptualization, T; Methodology, T, D.D.S; Investigation, D.I.A, M.S.N; Writing – Original Draft, D.I.A., T; Writing – Review & Editing, T, M.S.N; Funding Acquisition, T; Resources, D.D.S; Supervision, T, M.S.N.

Declaration of interest

None.

Received: September 1, 2022 Revised: December 26, 2022 Accepted: December 28, 2023

Published online: December 31, 2023

References

- Kemenkes. Buku Saku Hasil Studi Status Gizi Indonesia (SSGI) Tingkat Nasional, Provinsi, dan Kabupaten/Kota Tahun 2021. Kementerian Kesehatan Republik Indonesia, 2021 Des.
- Masoud MA, Kotb AS, Abd El-Raouf OM, Fikry EM.
 The neuroprotective effects of natural antioxidant against brain injury induced by paracetamol in a rat model of protein malnutrition. Egyptian Pharmaceutical Journal. 2020 Jan 1;19(1):55. https://doi.org/10.4103/epj.epj_54_19
- Gourine H, Grar H, Dib W, Mehedi N, Boualga A, Saidi D, Kheroua O. Effect of a normal protein diet on oxidative stress and organ damage in malnourished rats. Frontiers in Biology. 2018 Oct;13(5):366-75. https://doi.org/10.1007/ s11515-018-1511-5
- Alkadi H. A review on free radicals and antioxidants. Infectious Disorders-Drug Targets (Formerly Current Drug Targets-Infectious Disorders). 2020 Feb 1;20(1):16-26. https://doi.org/10.2174/18715265186661806281 24323
- Khare M, Mohanty C, Das BK, Jyoti A, Mukhopadhyay B, Mishra SP. Free radicals and antioxidant status in protein energy malnutrition. International journal of pediatrics. 2014 Oct;2014. https://doi.org/10.1155/2014/254396
- Gavia-García G, Rosas-Trejo MD, García-Mendoza E, Toledo-Pérez R, Königsberg M, Nájera-Medina O, Luna-Lopez A, González-Torres MC. t-BHQ protects against oxidative damage and maintains the antioxidant response in malnourished rats. Dose-response. 2018 Sep 25;16(3):1559325818796304. https://doi.org/10.1177/1559325818796304

- Stephensen CB. Primer on immune response and interface with malnutrition. InNutrition and infectious diseases 2021 (pp. 83-110). Humana, Cham. https:// doi.org/10.1007/978-3-030-56913-6_3
- Sanyoto DD, Noor MS. The effect of ethanol extract of pasak bumi (Eurycoma longifolia Jack.) on neurogenesis and neuroinflammation of rat post protein malnutrition. InIOP Conference Series: Earth and Environmental Science 2021 Nov 1 (Vol. 913, No. 1, p. 012091). IOP Publishing. https://doi.org/10.1088/1755-1315/913/1/012091
- De Aquino CC, Leitão RA, Oliveira Alves LA, Coelho-Santos V, Guerrant RL, Ribeiro CF, Malva JO, Silva AP, Oriá RB. Effect of hypoproteic and high-fat diets on hippocampal blood-brain barrier permeability and oxidative stress. Frontiers in nutrition. 2019 Jan 9;5:131. https://doi. org/10.3389/fnut.2018.00131
- Adelantado-Renau M, Beltran-Valls MR, Moliner-Urdiales D. Inflammation and cognition in children and adolescents: a call for action. Frontiers in Pediatrics. 2020 Sep 9;8:583. https://doi.org/10.3389/fped.2020.00583
- Goudet SM, Bogin BA, Madise NJ, Griffiths PL. Nutritional interventions for preventing stunting in children (birth to 59 months) living in urban slums in low-and middleincome countries (LMIC). Cochrane Database of Systematic Reviews. 2019(6). https://doi.org/10.1002/14651858. CD011695.pub2
- Scott N, Delport D, Hainsworth S, Pearson R, Morgan C, Huang S, Akuoku JK, Piwoz E, Shekar M, Levin C, Toole M. Ending malnutrition in all its forms requires scaling up proven nutrition interventions and much more: a 129-country analysis. BMC medicine. 2020 Dec;18(1):1-9. https://doi.org/10.1186/s12916-020-01786-5
- 13. Triawanti, Yunanto A, Sanyoto DD, Nur'amin HW. Nutritional Status Improvement in Malnourished Rat (Rattus norvegicus) after Seluang Fish (Rasbora spp.) Treatment. Current Research in Nutrition and Food Science Journal. 2018 Apr 20;6(1):127-34. https://doi.org/10.12944/CRNFSJ.6.1.14
- Triawanti, Sanyoto DD, Nur'amin HW. Reduction of Oxidative Stress by Seluang Fish (Rasbora spp.) in Brain of Malnourished Rats (Rattus norvegicus). International Journal of Food Engineering. 2018. 3(2): 107-111. https:// doi.org/10.18178/ijfe.3.2.107-111
- 15. Yunanto A, Didik DS, Triawanti, Meitria SN. 2014. Benefit of seluang fish (rasbora spp.)'S south kalimantan to the improvement of spatial memory quality. The 3rd International Symposium on wetlands environmental Management, Banjarmasin 8-9 Nopember 2014.
- 16. Hoorweg JC. Protein-energy malnutrition and intellectual abilities. InProtein-energy malnutrition and intellectual abilities 2019 Jul 8. De Gruyter Mouton.
- 17. Chaingam J, Choonong R, Juengwatanatrakul T, Kanchanapoom T, Putalun W, Yusakul G. Evaluation of anti-inflammatory properties of Eurycoma longifolia Jack

- and Eurycoma harmandiana Pierre in vitro cultures and their constituents. Food and Agricultural Immunology. 2022 Dec 31;33(1):530-45. https://doi.org/10.1080/09540105.2022.2100324
- Bai R, Yao C, Zhong Z, Ge J, Bai Z, Ye X, Xie T, Xie Y. Discovery of natural anti-inflammatory alkaloids: Potential leads for the drug discovery for the treatment of inflammation. European Journal of Medicinal Chemistry. 2021 Mar 5;213:113165. https://doi.org/10.1016/j.ejmech.2021.113165
- Triawanti, Sanyoto DD, Noor MS. The supplementation of pasak bumi (Eurycoma longifolia Jack.) in undernourished rats to increase spatial memory through antioxidant mechanism. Clinical Nutrition Experimental. 2020 Oct 1;33:49-59. https://doi.org/10.1016/j.yclnex.2020.08.002
- Roe K. An inflammation classification system using cytokine parameters. Scandinavian journal of immunology. 2021 Feb;93(2):e12970. https://doi.org/10.1111/sji.12970
- 21. Wu H, Denna TH, Storkersen JN, Gerriets VA. Beyond a neurotransmitter: The role of serotonin in inflammation and immunity. Pharmacological Research. 2019 Feb 1;140:100-14. https://doi.org/10.1016/j.phrs.2018.06.015
- 22. Sanyoto DD, Noor MS, Triawanti T. Potential Combinations of Pasak Bumi (Eurycoma longifolia Jack), Docosahexaenoic Acid, and Seluang Fish (Rasbora spp.) to Improving Oxidative Stress of Rats (Rattus norvegicus) Brain Undernutrition. Open Access Macedonian Journal of Medical Sciences. 2022 Jan 1;10(A):25-32. https://doi.org/10.3889/oamjms.2022.7671
- Sanyoto DD, Noor MS. The effect of ethanol extract of pasak bumi (Eurycoma longifolia Jack.) on neurogenesis and neuroinflammation of rat post protein malnutrition. InIOP Conference Series: Earth and Environmental Science 2021 Nov 1 (Vol. 913, No. 1, p. 012091). IOP Publishing. https://doi.org/10.1088/1755-1315/913/1/012091
- 24. Ruan J, Li Z, Zhang Y, Chen Y, Liu M, Han L, Zhang Y, Wang T. Bioactive constituents from the roots of Eurycoma longifolia. Molecules. 2019 Aug 30;24(17):3157. https://doi.org/10.3390/molecules24173157
- 25. Hien DT, Long TP, Thao TP, Lee JH, Trang DT, Minh NT, Van Cuong P, Dang NH, Dat NT. Anti-inflammatory effects of alkaloid enriched extract from roots of Eurycoma longifolia Jack. Asian Pacific Journal of Tropical Biomedicine. 2019 Jan 1;9(1):18. https://doi.org/10.4103/2221-1691.250265
- 26. Emelda E. Potensi Tongkat Ali (Eurycoma longifolia Jack.) Sebagai Anti Inflamasi. Jcps (Journal of Current Pharmaceutical Sciences). 2017 Sep 25;1(1):25-9.
- Calder PC. Marine ω-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids. 2015 Apr 1;1851(4):469-84. https://doi.org/10.1016/j.bbalip.2014.08.010
- 28. D'Angelo S, Motti ML, Meccariello R. ω -3 and ω -6 polyunsaturated fatty acids, obesity and cancer. Nutrients.

- 2020 Sep 10;12(9):2751. https://doi.org/10.3390/nu12092751
- 29. Muralidharan J, Papandreou C, Sala-Vila A, et al. Fatty Acids Composition of Blood Cell Membranes and Peripheral Inflammation in the PREDIMED Study: A Cross-Sectional Analysis. Nutrients. 2019;11(3):576. https://doi.org/10.3390/nu11030576
- 30. Yamaguchi A, Botta E, Holinstat M. Eicosanoids in inflammation in the blood and the vessel. Frontiers in Pharmacology. 2022:3973. https://doi.org/10.3389/fphar.2022.997403
- 31. Innes JK, Calder PC. Ω-6 fatty acids and inflammation. Prostaglandins Leukot Essent Fatty Acids. 2018;132:41-48. https://doi.org/10.1016/j.plefa.2018.03.004
- 32. Calder PC. Eicosapentaenoic and docosahexaenoic acid derived specialised pro-resolving mediators: Concentrations in humans and the effects of age, sex, disease and increased ω -3 fatty acid intake. Biochimie. 2020 Nov 1;178:105-23. https://doi.org/10.1016/j. biochi.2020.08.015
- 33. Tułowiecka N, Kotlęga D, Prowans P, Szczuko M. The role of resolvins: EPA and DHA derivatives can be useful in the prevention and treatment of ischemic stroke. International Journal of Molecular Sciences. 2020 Oct 15;21(20):7628. https://doi.org/10.3390/ijms21207628
- 34. Miao LH, Remø SC, Espe M, Philip AJ, Hamre K, Fjelldal PG, Skjærven K, Holen E, Vikeså V, Sissener NH. Dietary plant oil supplemented with arachidonic acid and eicosapentaenoic acid affects the fatty acid composition and eicosanoid metabolism of Atlantic salmon (Salmo salar L.) during smoltification. Fish & Shellfish Immunology. 2022 Apr 1;123:194-206. https://doi.org/10.1016/j.fsi.2022.02.049
- Che H, Li H, Song L, Dong X, Yang X, Zhang T, Wang Y, Xie W. Orally Administered DHA-Enriched Phospholipids and DHA-Enriched Triglyceride Relieve Oxidative Stress, Improve Intestinal Barrier, Modulate Inflammatory Cytokine and Gut Microbiota, and Meliorate Inflammatory Responses in the Brain in Dextran Sodium Sulfate Induced Colitis in Mice. Molecular Nutrition & Food Research. 2021 Aug;65(15):2000986. https://doi.org/10.1002/mnfr.202000986
- 36. Shibabaw T. Ω-3 polyunsaturated fatty acids: anti-inflammatory and anti-hypertriglyceridemia mechanisms in cardiovascular disease. Molecular and Cellular Biochemistry. 2021 Feb;476(2):993-1003. https://doi.org/10.1007/s11010-020-03965-7
- Calder PC. Ω-3 polyunsaturated fatty acids and inflammatory processes: nutrition or pharmacology?. British journal of clinical pharmacology. 2013 Mar;75(3):645-62. https:// doi.org/10.1111/j.1365-2125.2012.04374.x
- 38. Abou-Saleh H, Ouhtit A, Halade GV, Rahman MM. Bone benefits of fish oil supplementation depend on its EPA and DHA content. Nutrients. 2019 Nov 8;11(11):2701. https://doi.org/10.3390/nu11112701

- 39. Sogandi S, Sanjaya RE, Baity N, Syahmani S. Identifikasi kandungan gizi dan profil asam amino dari ikan seluang [rasbora sp] (identification of nutritional content and profiles of amino acid from seluang fish [Rasbora Sp]). Penelitian Gizi dan Makanan (The Journal of Nutrition and Food Research). 2019;42(2):73-80. https://doi.org/10.22435/pgm.v42i2.1287
- He F, Wu C, Li P, Li N, Zhang D, Zhu Q, Ren W, Peng Y. Functions and signaling pathways of amino acids in intestinal inflammation. BioMed research international. 2018 Oct;2018. https://doi.org/10.1155/2018/9171905
- 41. Bae JY, Koo GH, Park SC, Shin KO. Effects of branchedchain amino acid and glutamine supplementation on angiogenic factors and pro-inflammatory cytokines after acute exercise in adolescence athletes. The Asian Journal of Kinesiology. 2019 Apr 30;21(2):51-8. https://doi. org/10.15758/ajk.2019.21.2.51
- 42. Farré R, Fiorani M, Abdu Rahiman S, Matteoli G. Intestinal permeability, inflammation and the role of nutrients. Nutrients. 2020 Apr 23;12(4):1185. https://doi.org/10.3390/nu12041185
- 43. Sugihara K, Morhardt TL, Kamada N. The role of dietary nutrients in inflammatory bowel disease. Frontiers in Immunology. 2019 Jan 15;9:3183. https://doi.org/10.3389/fimmu.2018.03183
- 44. Sorgdrager FJ, Naudé PJ, Kema IP, Nollen EA, Deyn PP. Tryptophan metabolism in inflammaging: from biomarker to therapeutic target. Frontiers in Immunology. 2019 Oct 30;10:2565. https://doi.org/10.3389/fimmu.2019.02565
- 45. Hou YC, Wu JM, Wang MY, Wu MH, Chen KY, Yeh SL, Lin MT. Glutamine supplementation attenuates expressions of adhesion molecules and chemokine receptors on T cells in a murine model of acute colitis. Mediators of Inflammation. 2014 Oct;2014. https://doi.org/10.1155/2014/837107
- 46. Fazio F, Ulivieri M, Volpi C, Gargaro M, Fallarino F. Targeting metabotropic glutamate receptors for the treatment of neuroinflammation. Current Opinion in Pharmacology. 2018 Feb 1;38:16-23. https://doi.org/10.1016/j.coph.2018.01.010
- 47. Hasegawa T, Mizugaki A, Inoue Y, Kato H, Murakami H. Cystine reduces tight junction permeability and intestinal inflammation induced by oxidative stress in Caco-2 cells. Amino Acids. 2021 Jul;53(7):1021-32. https://doi.org/10.1007/s00726-021-03001-y
- 48. Ting LE, Leach ST, Lemberg DA, Day AS. A Brief Overview of Nutrient Anti-Inflammatory Molecules and their In Vitro and In Vivo Activity. Journal of Nutritional Medicine and Diet Care. 2016;2(2). https://doi.org/10.23937/2572-3278.1510018
- 49. Wu T, Wang C, Ding L, Shen Y, Cui H, Wang M, Wang H. Arginine relieves the inflammatory response and enhances the casein expression in bovine mammary epithelial cells induced by lipopolysaccharide. Mediators of Inflammation. 2016 Jan 1;2016. https://doi.org/10.1155/2016/9618795

- Bao X, Feng Z, Yao J, Li T, Yin Y. Roles of dietary amino acids and their metabolites in pathogenesis of inflammatory bowel disease. Mediators of inflammation. 2017 Oct;2017. https://doi.org/10.1155/2017/6869259
- 51. Wessler LB, de Miranda Ramos V, Pasquali MA, Moreira JC, de Oliveira J, Scaini G, Streck EL. Administration of branched-chain amino acids increases the susceptibility to lipopolysaccharide-induced inflammation in young Wistar rats. International Journal of Developmental Neuroscience. 2019 Nov 1;78:210-4. https://doi.org/10.1016/j.ijdevneu.2019.07.007
- Kato H, Miura K, Nakano S, Suzuki K, Bannai M, Inoue Y. Leucine-enriched essential amino acids attenuate inflammation in rat muscle and enhance muscle repair after eccentric contraction. Amino Acids. 2016 Sep;48(9):2145-55. https://doi.org/10.1007/s00726-016-2240-1
- 53. Da Silva MS, Bigo C, Barbier O, Rudkowska I. Whey protein hydrolysate and branched-chain amino acids downregulate inflammation-related genes in vascular endothelial cells. Nutrition Research. 2017 Feb 1;38:43-51. https://doi.org/10.1016/j.nutres.2017.01.005
- Holeček M. Branched-chain amino acids in health and disease: metabolism, alterations in blood plasma, and as supplements. Nutrition & metabolism. 2018 Dec;15(1):1-2. https://doi.org/10.1186/s12986-018-0271-1
- 55. Cooper AJ. Glutamine synthetase. InGlutamine and glutamate in mammals 2018 Jan 18 (pp. 7-32). CRC Press. https://doi.org/10.1201/9781351072298-3
- Shu XL, Yu TT, Kang K, Zhao J. Effects of glutamine on markers of intestinal inflammatory response and mucosal permeability in abdominal surgery patients: A meta-analysis. Experimental and therapeutic medicine. 2016 Dec 1;12(6):3499-506. https://doi.org/10.3892/ etm.2016.3799
- 57. Kim MH, Kim H. The roles of glutamine in the intestine and its implication in intestinal diseases. International journal of molecular sciences. 2017 May 12;18(5):1051. https://doi.org/10.3390/ijms18051051
- Cruzat V, Macedo Rogero M, Noel Keane K, Curi R, Newsholme P. Glutamine: metabolism and immune function, supplementation and clinical translation. Nutrients. 2018 Oct 23;10(11):1564. https://doi. org/10.3390/nu10111564
- 59. Yan S, Hui Y, Li J, Xu X, Li Q, Wei H. Glutamine relieves oxidative stress through PI3K/Akt signaling pathway in DSS-induced ulcerative colitis mice. Iranian Journal of Basic Medical Sciences. 2020 Sep;23(9):1124.
- Chen L, Deng H, Cui H, et al. Inflammatory responses and inflammation-associated diseases in organs. Oncotarget. 2017;9(6):7204-7218. https://doi.org/10.18632/oncotarget. 23208
- 61. Luo LL, Li YF, Shan HM, Wang LP, Yuan F, Ma YY, Li WL, He TT, Wang YY, Qu MJ, Liang HB. L-glutamine protects mouse brain from ischemic injury via up-

- regulating heat shock protein 70. CNS neuroscience & therapeutics. 2019 Sep;25(9):1030-41. https://doi.org/10.1111/cns.13184
- 62. Petry ÉR, de Freitas Dresch D, Carvalho C, Medeiros PC, Rosa TG, de Oliveira CM, Martins LA, Schemitt E, Bona S, Guma FC, Marroni NP. Oral glutamine supplementation attenuates inflammation and oxidative stress-mediated skeletal muscle protein content degradation in immobilized rats: Role of 70 kDa heat shock protein. Free Radical Biology and Medicine. 2019 Dec 1;145:87-102. https://doi.org/10.1016/j.freeradbiomed.2019.08.033
- 63. Deters BJ, Saleem M. The role of glutamine in supporting gut health and neuropsychiatric factors. Food Science and Human Wellness. 2021 Mar 1;10(2):149-54. https://doi.org/10.1016/j.fshw.2021.02.003
- 64. Manuel Apolinar L, Rocha L, Datriawamasio L, Tesoro Cruz E, Zarate A. Role of prenatal undernutrition in the expression of serotonin, dopamine and leptin receptors in adult mice: implications of food intake. Molecular medicine reports. 2014 Feb 1;9(2):407-12. https://doi.org/10.3892/mmr.2013.1853
- 65. Mokler DJ, McGaughy JA, Bass D, Morgane PJ, Rosene DL, Amaral AC, Rushmore RJ, Galler JR. Prenatal protein malnutrition leads to hemispheric differences in the extracellular concentrations of norepinephrine, dopamine and serotonin in the medial prefrontal cortex of adult rats. Frontiers in Neuroscience. 2019 Mar 5;13:136. https://doi.org/10.3389/fnins.2019.00136
- 66. Hernández-Rodríguez J, Mondragón-Herrera JA, Boyzo-Montes de Oca A, Mercado-Camargo R, & Manjarrez-Gutiérrez G. How intrauterine growth restriction due to nutritional stress changes the function of key proteins in brain serotonin metabolism during development. Boletín médico del Hospital Infantil de México. 2021; 78(6), 571-583. https://doi.org/10.24875/BMHIM.20000334
- 67. Friedman M. Analysis, nutrition, and health benefits of tryptophan. International Journal of Tryptophan Research. 2018; 11, 1178646918802282. https://doi.org/10.1177/1178646918802282
- 68. Manjarrez-Gutiérrez G, Hernández-Rodríguez J, & Mondragón-Herrera JA. Nutritional Recovery and its Effect on Tryptophan-5-Hydroxylases Expression. Cell Number and on Changes Caused by Intrauterine Growth Restriction in the Developing Brain. J Nutr Food Sci. 2020; 10, 774
- Van Galen KA, Ter Horst KW, & Serlie MJ. Serotonin, food intake, and obesity. Obesity Reviews. 2021; 22(7), e13210. https://doi.org/10.1111/obr.13210
- Siotto M, Germanotta M, Santoro M, et al. Serotonin Levels and Cognitive Recovery in Patients with Subacute Stroke after Rehabilitation Treatment. Brain Sci. 2021;11(5):642. https://doi.org/10.3390/brainsci11050642
- 71. Stahl SM. Beyond the dopamine hypothesis of schizophrenia to three neural networks of psychosis: dopamine, serotonin,

- and glutamate. CNS spectrums. 2018 Jun;23(3):187-91. https://doi.org/10.1017/S1092852918001013
- 72. Haleem DJ, & Mahmood K. Brain serotonin in high-fat diet-induced weight gain, anxiety and spatial memory in rats. Nutritional neuroscience. 2021; 24(3), 226-235. https://doi.org/10.1080/1028415X.2019.1619983
- 73. Daray FM, Mann JJ, Sublette ME. How lipids may affect risk for suicidal behavior. J Psychiatr Res. 2018;104:16-23. https://doi.org/10.1016/j.jpsychires.2018.06.007
- 74. Lange KW. Ω -3 fatty acids and mental health. Global Health Journal. 2020 Mar 1;4(1):18-30. https://doi.org/10.1016/j.glohj.2020.01.004